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Core-binding Factor Acute Myeloid Leukemia with Inv(16): Older Age and High White Blood Cell Count are Risk Factors for Treatment Failure

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LETTER TO THE EDITOR

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Dear Editors:

The core-binding factor (CBF) and acute myeloid leukemias (AML) are considered a distinct subgroup of AML due to similar molecular pathogenesis and clinical features.[1] The two forms of CBF-AML have rearrangements t(8;21)(q22;q22.1) [t(8;21)] and inv(16) (p13.1q22) or t(16;16)(p13.1;q22) [inv(16)] which result in the fusion genes *RUNX1/RUNXT1* and *CBFB/MYH11*, respectively. Although survival of CBF-AML is superior than other subtypes of AML, relapse occurs in approximately 30%-40% of patients.[1-4] Finding risk factors for treatment failure (eg, relapse or death) may be useful to manage patients individually. However, the relative rarity of CBF-AML (approximately 15%-20% of AML cases) and its relatively good prognosis has confounded these efforts.[2,3] Moreover, recent studies clearly indicate each CBF-AML is a distinct disease regarding patient and disease characteristics,[2-5] and should be evaluated separately. We previously published a scoring system (I-CBFit) for treatment failure for t(8;21),[6] now here we present our analysis for inv(16).

Eleven US and Europe centers collected data on 550 patients with CBF-AML; 290 patients had inv(16). Inclusion criteria were as follows: (a) AML patients with inv(16), t(16;16), or *CBFB/MYH11* by fluorescence in situ hybridization confirmed by the reporting institutions; (b) cases diagnosed between July 1996 and January 2017, (c) diagnostic or at least postinduction bone marrow sample was available for review in the participating institution's pathology department. Data were uniformly collected using a predesigned data spreadsheet. The data forms included the following: patient characteristics (age, sex, and race); disease characteristics [date of diagnosis, white blood cell count (WBC) at diagnosis ($\times10^9$/L), cytogenetics, *KIT* D816V mutational status, primary or secondary AML]; hematopoietic cell transplantation (HCT) (autologous HCT or allogeneic HCT (alloHCT), donor type, remission status at HCT); and events (relapse, death, or alive at last contact). Deidentified patient data were transferred to the University of Minnesota where the main database was created and managed. This study was approved by the Institutional Review Board: Human Subjects Committee at the University of Minnesota.

Secondary AML was assigned if a patient had a history of chemotherapy/radiation therapy for a malignancy and/or had a history of preleukemic neoplastic bone marrow disease. The presence of 46 chromosomes in one clone or each clone was defined

TABLE 1 Characteristics of patients with inv(16) achieving CR

Variable	Total
n	264
Age (median [range])	49 [5-78]
Sex, n (%)	
Female	129 (48.9)
Male	117 (44.3)
Missing	18 (6.8)
WBC at Diagnosis (10^9/L) (median [range])	21.30 [1-373]
Missing n (%)	5 (1.9)
AML type, n (%)	
Primary	219 (83.0)
Secondary	32 (12.1)
Missing	13 (4.9)
Chromosome 4 abnormalities, n (%)	
No	243 (92.0)
Yes	5 (1.9)
Missing	16 (6.1)
Chromosome 5 or 7 abnormalities, n (%)	
No	233 (88.3)
Yes	15 (5.7)
Missing	16 (6.1)
+8, n (%)	
No	212 (80.3)
Yes	36 (13.6)
Missing	16 (6.1)
Chromosome 9 abnormalities, n (%)	
No	243 (92.0)
Yes	5 (1.9)
Missing	16 (6.1)
+ 22, n (%)	
No	208 (78.8)
Yes	40 (15.2)
Missing	16 (6.1)

(Continues)

TABLE 1 (Continued)

Variable	Total
-X,n (%)	
No	245 (92.8)
Yes	3 (1.1)
Missing	16 (6.1)
-Y,n (%)	
No	239 (90.5)
Yes	9 (3.4)
Missing	16 (6.1)
Number of chromosomes, n (%)	
<46	6 (2.3)
=46	180 (68.2)
>46	62 (23.5)
Missing	16 (61)
KIT D816V mutation, n (%)	
Negative	135 (51.1)
Positive	29 (11.0)
Missing	100 (37.9)
AlloHCT, n (%)	92 (34.8)
Disease status at alloHCT, n (%)	
No CR	7 (7.7)
CR1	28 (30.8)
CR2	54 (59.3)
>CR2	1 (1)
Missing	2 (2)
Relapsed within 2 y (%) in CR1	91 (34.5)
Nonrelapse mortality within 2 y in CR1, n (%)	15 (5.7)
EFS duration, months (median)	25.5
DFS duration, months (median)	29.5
OS duration, months (median)	Not reached

Abbreviations: AlloHCT, allogeneic hematopoietic cell transplantation; CR, complete remission; DFS, disease-free survival; EFS, event-free survival; OS, overall survival; WBC, white blood cell count.

as pseudodiploidy (due to inversion or t(16;16) it was not named diploidy), and if the chromosome number was higher or lower than 46 chromosomes in any clone (in patients with more than one clone) it was defined as hyperdiploidy and hypodiploidy, respectively. Event-free survival (EFS) was defined as the time from diagnosis to relapse, death from any cause, or failure to achieve complete remission. Disease-free survival (DFS) was defined as the time from complete remission to relapse or death from any cause. Overall survival (OS) was defined as the time from diagnosis to last follow-up alive or date of death of any cause. We described patients' characteristics using the median and range for continuous variables, and frequency and percentage for categorical variables. All patients were included EFS and OS analyses, but subjects who did not achieve complete remission (CR) were excluded from DFS analysis. Subjects that were censored before 2 years of follow-up were considered to be missing for the primary outcome. Covariates considered for inclusion were sex, age, race, primary or secondary AML, WBC at diagnosis, + 22, +8, and diploid status. Other chromosome abnormalities (4, 5, 7, 9, X, Y) were not considered for the prognostic model due to their rarity in our sample. KIT mutations were not included due to too many with missing data. Missing data were imputed with multivariate Imputation by Chained Equations in R. Twenty-five imputed datasets were generated. Stepwise model selection based on Akaike Information Criterion was performed on each imputed dataset. Covariates that were selected in at least half of the imputed datasets were included in the final model. Parameter estimates and standard errors were computed according to Rubin's rules. A Cox proportional hazards model was initially used to investigate the effects of age, sex, WBC at diagnosis, +8, +22, and hyperdiploidy on EFS. An event was defined as either relapse, death, or treatment failure (defined as not achieving CR at 30 days after diagnosis). Patients were considered to be censored if they were lost to follow-up or received alloHCT in CR1. When it was found that age and WBC were the only significant predictors, cutoff values were determined to group patients into high- and low-risk categories. Models for DFS and OS were also
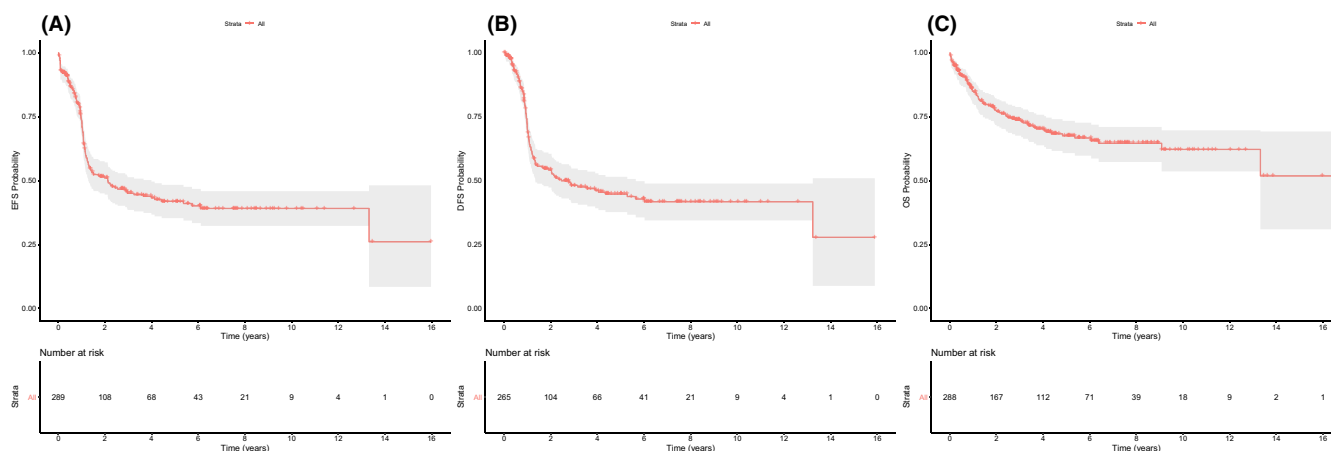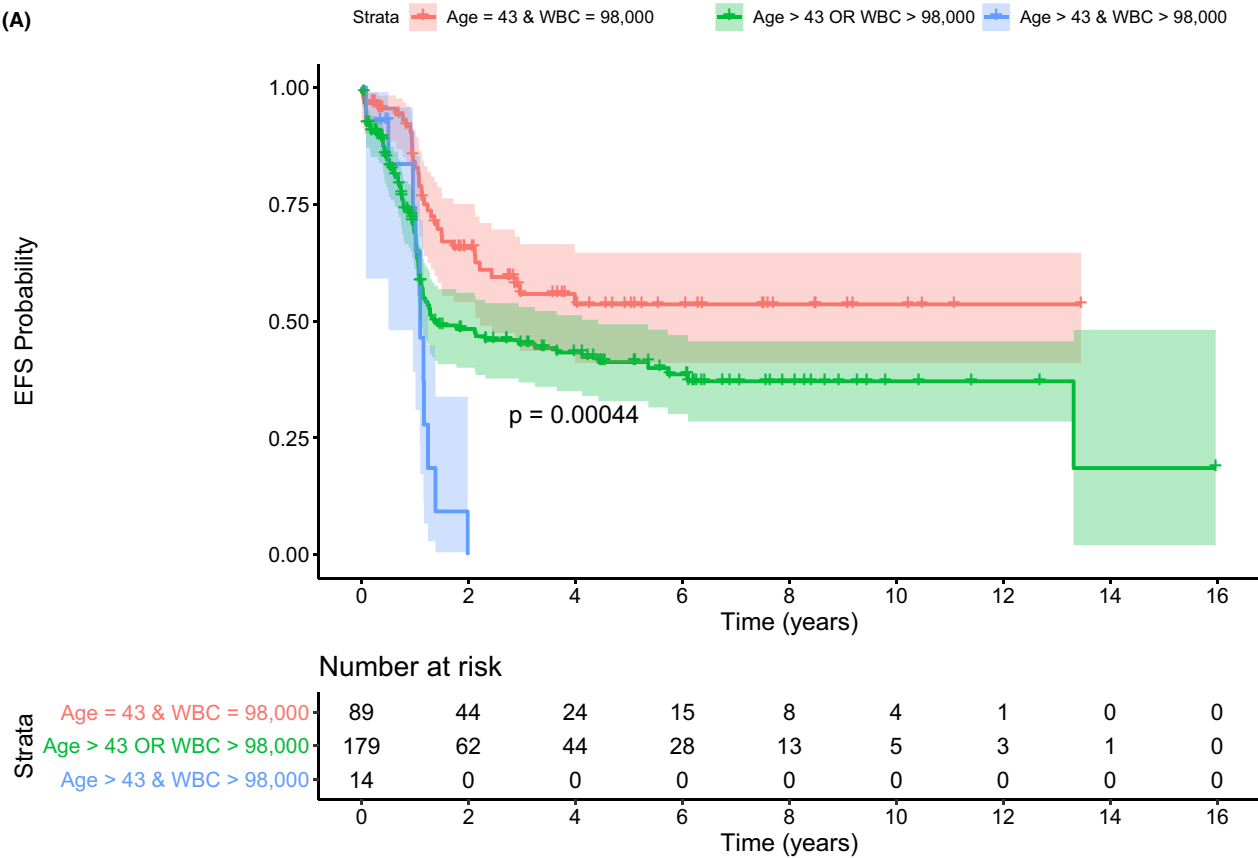

FIGURE 1 EFS (A), DFS (B), and OS (C) in patients with inv(16) [Colour figure can be viewed at wileyonlinelibrary.com]

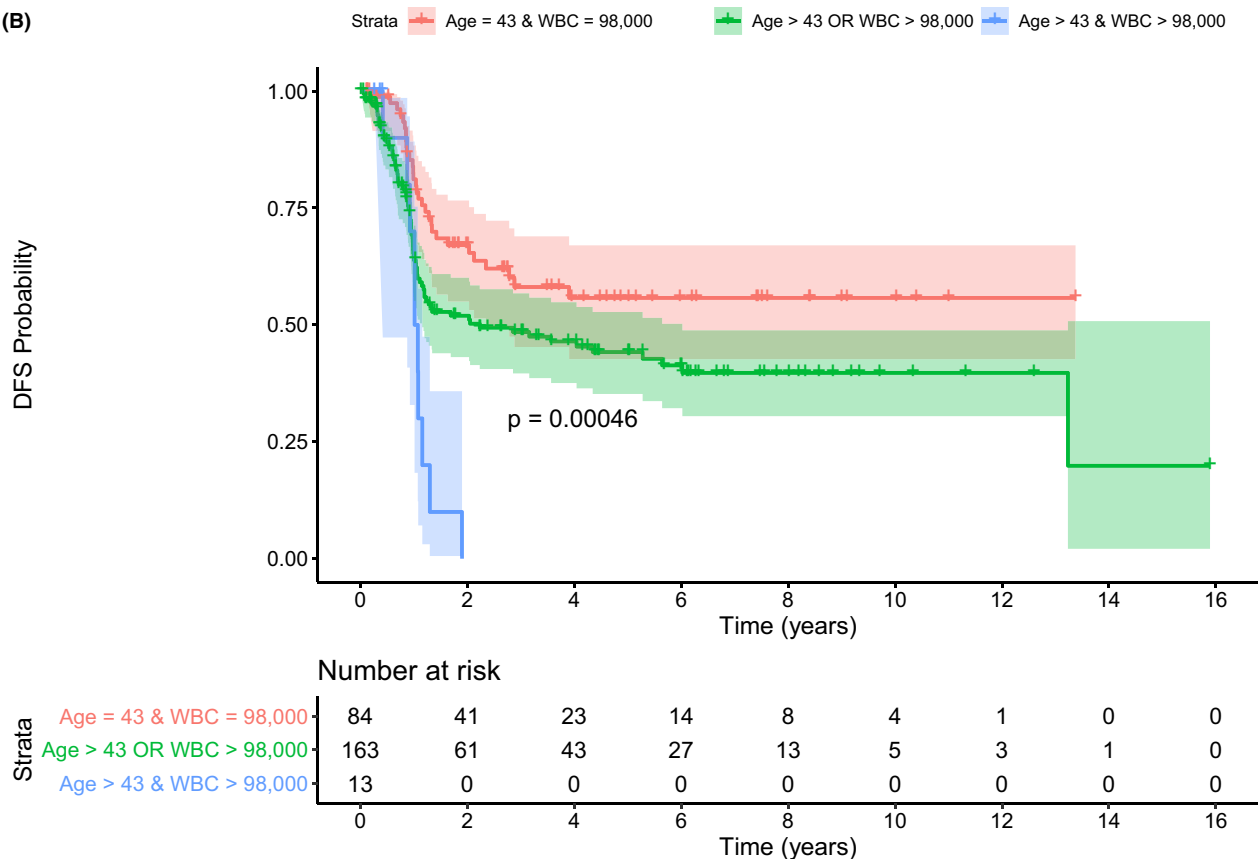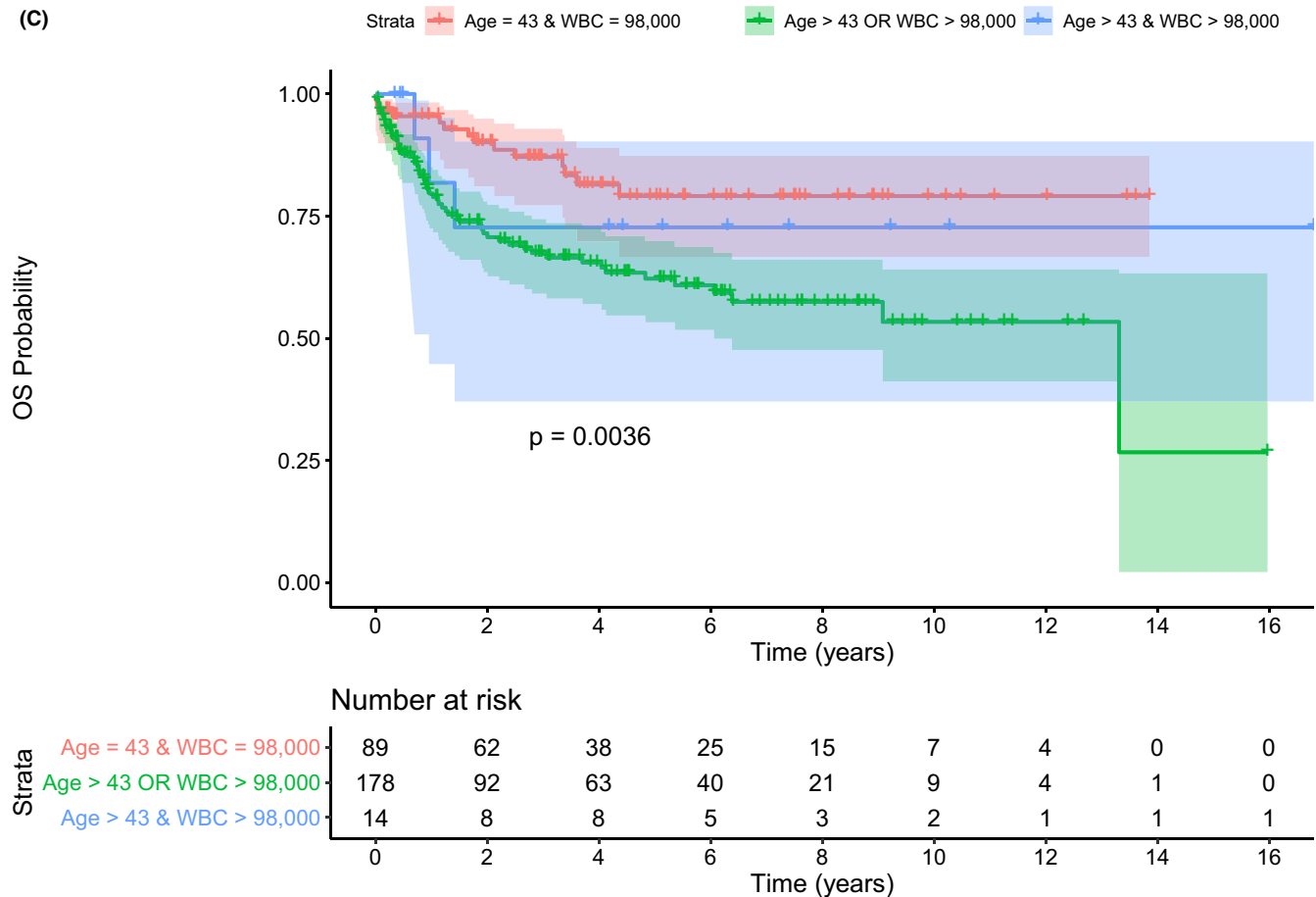**(A)**

(B)

FIGURE 2 (Continued)

(C)

| Strata | Age = 43 & WBC = 98,000 | Age > 43 OR WBC > 98,000 | Age > 43 & WBC > 98,000 |

Number at risk

Strata	0	2	4	6	8	10	12	14	16
Age = 43 & WBC = 98,000	89	62	38	25	15	7	4	0	0
Age > 43 OR WBC > 98,000	178	92	63	40	21	9	4	1	0
Age > 43 & WBC > 98,000	14	8	8	5	3	2	1	1	1

FIGURE 2 EFS (A), DFS (B), and OS (C) were shown among patients with no (age ≤43 y and WBC ≤ 98 × 10^9/L), 1 (age ≤ 43 y or WBC ≤ 98 × 10^9/L) and 2 risk factors (age >43 y and WBC > 98 × 10^9/L)

computed. Separately for age and WBC, a ROC curve was generated for EFS for generated at 2 years after diagnosis using the survivalROC package in R. Cutoff points were chosen to maximize the Youden Index (Specificity + Sensitivity – 1). Kaplan-Meier curves and log-rank test were used for survival analyses in patients who had known diagnostic WBC and age (n = 282).

Of the 290 patients with inv16 (median age of 49 years), 264 achieved CR1 (Table 1). The most common additional cytogenetic abnormalities were +8 and +22. Hyperdiploidy (23.5%) was more frequent. *KIT* D816V mutation was present in 17.7% of tested patients.

Relapse or death occurred in 106 patients (40.2%) within 2 years after CR1 at a median of 10.7 months (range 0.8 to 22.8 months). The median EFS and DFS were 25.5 and 29.5 months, respectively while OS was not reached. (Table 1 and Figure 1). In multivariate analysis (MVA) for survival at 2-year, older age (≥43 years) was associated with inferior EFS [(Hazard ratio (HR): 1.017, 95% Confidence Interval (CI): (1.007-1.028), P = .0014], DFS [HR: 1.014, 95% CI (1.003-1.026), P = .0123], and OS [HR: 1.034, 95% CI (1.018-1.049), P = .00002]. Higher white blood cell counts (WBC ≥ 98 × 10^9/L) were associated with a poorer EFS [HR: 1.005, 95% CI (1.002-1.008), P = .0003] and DFS [HR: 1.0006, 95% CI (1.004-1.009), P = .0001], but not with OS [HR: 1000 95% CI (0.995-1.005), P = .958].

Patients stratified by age and WBC into three groups (age ≤43 years and WBC ≤98 × 10^9/L vs age ≤43 years or WBC ≤98 × 10^9/L vs age >43 years and WBC >98 × 10^9/L). Regarding EFS, younger patients with a lower WBC (no risk group, age ≤43 years and WBC ≤98 × 10^9/L) did not reach median survival whereas those with 1 risk (age ≤43 years or WBC ≤98 × 10^9/L) and patients with 2 risks (age >43 years and WBC > 98 × 10^9/L) had a median of EFS of 16.3 months and 13.1 months, respectively, P = .00044 (Figure 2A). Regarding DFS, patients with no risk did not reach median survival whereas those with 1 risk and those 2 risks (age >43 years and WBC >98 × 10^9/L) had a median DFS of 26.8 months and 12.5 months, respectively, P = .00046 (Figure 2B). Regarding OS, patients with no risk or with both risks did not reach median survival whereas those with 1 risk had a median OS of 160 months, P = .0036 (Figure 2C).

Although this multicenter study had limitations (due to retrospective in nature and thus missing detailed molecular data and no data on minimal residual disease), the data from a large number of patients with long-term follow-up allowed us to identify clear risk factors for treatment failure; older patients and higher WBC counts. Hoyos et al also showed that higher WBC count (> 20 × 10^9/L) had higher relapse at 5-year (40% vs

16%, $P = .001$) in 150 patients with CBF-AML (76 with CBFB-MYH11).[7] When patients were stratified by age and WBC (age >50 years and WBC > 20×10^9/L), similar to our results, patients who had no risk factor had a superior 5-year OS (80%). Other studies have also supported that WBC (one defined it similar to ours, 100×10^9/L) is associated with poor DFS, mainly due to increased relapse..[8,9] Pashka et al reported that patients with higher WBC had decreased DFS (HR = 1.33, $P = .02$) and more mutated *KIT* and *FLT3*.[8] De Jonge et al showed low WBC (<20×10^9/L) was associated with a higher CR rate ($P = .024$), improved EFS (median 77.2 vs 9.7 months) and improved OS (median 85.5 vs 28.9 months, $P = .001$) in favorable prognostic AML.[10] Delauney et al found that older aged (>36 years), +22, and severe thrombocytopenia (<30×10^9/L) were associated with poor DFS in 110 patients with inv(16).[11] In MVA, age was the only risk factor for poor DFS. The impact of +22 on survival is conflicting[2,3,8,11] and might be more prominent in patients with lower WBC.[3]

A *KIT* mutation (exons 8-13 & 17), present in up to 37% of patients, was associated with poor prognosis for survival in CBF-AML.[4,8] *KIT* mutations seem to have an impact in patients with t(8;21), but not in those with inv16.[6,12] However, in the current study of patients with inv16, *KIT* mutations could not be tested due to excessive missing data. Similar to our findings, complex karyotype[2,5,11] and secondary AML did not have effect in treatment failure.[13]

Prospective studies are warranted to improve survival in older inv(16) patients with a higher WBC at diagnosis.

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CONFLICT OF INTEREST

The authors have no conflict of interest relevant to the study to disclose.

AUTHOR CONTRIBUTIONS

CU conceived the idea, collected data, analyzed data, searched literature, wrote the manuscript, and edited the manuscript. ER and TB involved in execution of data analysis, writing and editing article. CA, PV, JG, HJD, and DW conceived the idea, writing and editing the manuscript. TG and H-PH involved in pathologic examination of BMs as needed, identifying patients, data collection, wrote the manuscript, and edited the manuscript. KM involved in cytogenetic confirmation, writing and editing the manuscript. EAM, SP, YK, RO, HV, CY, CAY, ML, SN, JLH, TKK, LS, LBB, MBM, WS, VP, NKK, DC, VP, AR, SK, GH, CDB, JK, GM, GB, CB, CC, JK, MRL, MAP, AS, RN, SYH, SBO, and A-IS involved in identifying patients, data collection, wrote the manuscript, and edited the manuscript.

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